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# **Genetic analysis of shoot regeneration from cotyledonary explants in** *Brassica napus*

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**Abstract** Genetic analysis of shoot regeneration from cotyledonary explants of Brassica napus was carried out by 7×7 diallel crosses using cultivars showing a different ability for regeneration. Both additive and dominant effects were significant, with the additive effect being more important than the dominant one. Dominant genes had a positive effect on shoot regeneration. Non-allelic interaction and average maternal effects were not detected, while specific the maternal one was significant. In the 5×5 sub-diallel table, the maternal effect became nonsignificant. The mean degree of dominance was 0.759. Broad- and narrow-sense heritabilities were 0.973 and 0.819, respectively, indicating that shoot regeneration ability can be easily transferred into economically important cultivars showing a low or an unresponsive ability.

**Key words** Diallel analysis · Regeneration ability · Cotyledonary culture · *Brassica napus* · Heritablity

## Introduction

Efficient plant regeneration via organogenesis or embryogenesis is indispensable to the application of tissue culture technology for plant improvement. However, great variation among genotypes in the ability of plant regeneration was observed in many plants. Such variation can be an obstacle to utilizing tissue culture for practical breeding. Genetic factors controlling the ability of tissue culture have been reported in several plants such as rice (Abe and Futsuhara 1991; Taguchi-Shiobara et al. 1997), barley (Komatsuda et al. 1989), maize, (Petolino and Thompson 1987) and wheat (Lazar et al. 1984). Re-

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cently, the loci involved in regeneration ability have been identified using molecular markers in several plants (Armstrong et al. 1992; Koornneef et al. 1993; Komatsuda et al. 1995; Yamagishi et al. 1996).

In *Brassica* crops various kinds of plant regeneration systems have been developed (Palmer and Keller 1994), but little work has been carried out on the genetic factors involved in the regeneration ability. Shoot regeneration systems using cotyledonary explants have been developed in some Brassicas and employed for genetic transformation, due to their high regeneration ability (Narasimhulu and Chopra 1988; Moloney et al. 1989; Hachey et al. 1991; Babic et al. 1998). We recently reported that the ability for shoot regeneration from cotyledonary explants was strongly influenced by genotype in rapeseed (Brassica napus) (Ono et al. 1994) and Chinese cabbage (Brassica campestris) (Zhang et al. 1998). In the present study, we have attempted to obtain genetic information on shoot regeneration from cotyledonary explants of *B. napus* by diallel analysis.

### **Materials and methods**

Seven cultivars of rapeseed (B. napus) (see Table 1), were chosen, based on the results of a previous study in which they showed different abilities for shoot regeneration from cotyledons (Ono et al. 1994). A diallel set of crosses was performed among the seven cultivars. The ability of shoot regeneration from cotyledonary explants was examined for 42 reciprocal crosses and seven selfed parents. Tissue culture conditions and medium composition were performed as previously described (Ono et al. 1994). Seeds were sterilized in sodium hypochlorite (1.0% active chlorite) with 1-2 drops of Tween-20 for 20 min. After rinsing in sterile distilled water three times, the seeds were placed in 15×90 mm Petri dishes containing growth regulator-free MS agar (0.7%) medium (Murashige and Skoog 1962) at density of 20 seeds per dish. Cotyledonary explants excised from 4-day old seedlings were embedded into the regeneration media which was that composed of MS medium supplemented with 4 mg/l of 6-benzylaminopurine. Ten explants were cultured in a 15×90 mm Petri dish. After 3 weeks of culture, the adventitious shoots formed on the explants were counted. The regeneration frequency (number of explants with shoots/total number of explants) was averaged for at least three replications.

Statistical analysis using either direct or arcsin-transformed data was performed according to the methods by Hayman (1954a, b) and Jinks (1954). Calculation in the diallel analysis was carried out using the computer program 'DIALL' developed by Ukai (1989). The narrow- and broad-sense heritabilities were calculated according to the method of Mather and Jinks (1971).

### **Results and discussion**

The frequency of shoot regeneration for the seven parents ranged from 0% to 96.7% (Table 1). 'Arabella', 'Doral' and 'Westar' showed a high frequency of regeneration, while 'Topas', 'Bridger' and 'Cascade' showed a low frequency and 'Norin 30' could regenerate no shoots. Among  $42 \, F_1$  combinations, the regeneration frequencies of many  $F_1$ s were similar to those of the high-responsive parent. The regeneration ability of less-

**Table 1** Shoot regeneration frequency (%) from cotyledon explants in seven parents and their  $F_1$  hybrids of *B. napus*. The underlined figures represent parental values

Female	Male						
	1	2	3	4	5	6	7
1 Arabella 2 Doral 3 Westar 4 Topas 5 Bridger 6 Cascade 7 Norin 30	96.7 96.7 76.7 53.3 83.0 83.3 93.3	90.0 93.3 70.0 46.7 86.7 78.5 73.3	86.7 43.3 86.7 70.0 33.3 30.0 70.0	73.3 78.5 50.0 13.3 36.7 13.3 6.7	90.0 78.5 43.3 20.0 <u>6.7</u> 6.7 33.3	83.3 86.7 43.3 6.7 3.3 <u>6.7</u> 0.0	93.3 80.0 40.0 20.0 16.7 6.7 0.0

**Table 2** Analysis of variance of a 7×7 diallel table for shoot regeneration, after Hayman (1954a)

Source	df	SS	MS	F
a b b1 b2 b3	6 21 1 6 14	39569.200 11155.700 600.095 1998.800 8556.760	6599.370 531.222 600.095 333.133 611.197	232.16** 18.69** 21.11** 11.72** 21.50**
c d Error	6 15 96	136.932 1815.060	22.822 121.004 28.426	0.80 4.26**

<sup>\*, \*\*</sup> Significant at the 5% and 1% levels, respectively

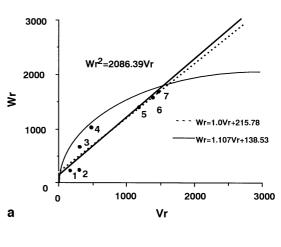
**Table 3** Analysis of variance of 5×5 sub-diallel table for shoot regeneration, after Hayman (1954a)

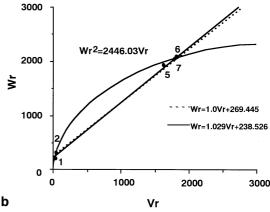
Source	df	SS	MS	F
a	4	29260.000	7314.990	320.07**
b	10	6896,980	689.698	30.18**
b1	1	2025.300	2025.300	88.62**
b2	4	321.871	80.468	3.52*
b3	5	4549.810	909.962	39.82**
c	4	54.972	13.743	0.60
d	6	247.024	41.171	1.80
Error	48		22.854	

<sup>\*, \*\*</sup> Significant at the 5% and 1% levels, respectively

responsive cultivars could be improved by crossing with high-responsive ones. Heterotic combinations were found in those between 'Bridger' and 'Topas' and between 'Bridger' and 'Norin 30'.

As two calculations for statistical analysis using direct and arcsin-transformed data gave the similar results, only the former are described in this paper. Analysis of variance of a diallel table for shoot regeneration revealed that both additive (a) and dominant (b) effects were significant (Table 2). This is consistent with the results obtained from genetic analysis of the regeneration ability of other crops such as rice (Peng and Hodes 1989; Abe and Futsuhara 1991; Taguchi-Shiobara et al. 1997), barley (Komatsuda et al. 1989) and maize (Petolino and Thompson 1987). The average maternal effect (c) was not significant, while the specific maternal one (d) was. The existence of maternal effects on regeneration ability were contrary to studies using other crops. This may be due to differences in the genotype employed. The relatively high maternal effects were detected in the combinations using 'Westar' and 'Topas' as a parent. Therefore, these two cultivars were eliminated from the diallel table to make a 5×5 sub-diallel table. In the analysis of the sub-diallel table, additive and dominant effects were significant, but the maternal one became nonsignificant (Table 3).





**Fig. 1** Vr, Wr graph for shoot regeneration from a 7×7 diallel table (**a**) and a 5×5 sub-diallel table (**b**). *I* 'Arabella', 2 'Doral', 3 'Westar', 4 'Topas', 5 'Bridger', 6 'Cascade', 7 'Norin 30'

**Table 4** Estimates of genetic parameters for shoot regeneration derived from 5×5 sub-diallel table

Component	Estimated value±SE
D	2461.690±67.851
F	417.148±158.912
H <sub>1</sub>	1440.510+154.597
$ H_2 $ $ h^2 $	1393.540±151.322 1299.170±162.243
E	22.854
(H1/D) <sup>1/2 a</sup>	0.759
H <sub>2</sub> /4H <sub>1</sub> <sup>b</sup>	0.383
{(4DH <sub>1</sub> ) <sup>1/2</sup> +F}{4DH <sub>1</sub> ) <sup>1/2</sup> -F} <sup>c</sup>	0.299
h <sup>2</sup> (ns) <sup>d</sup>	0.819
h <sup>2</sup> (bs) <sup>e</sup>	0.973

- a Mean dominance
- <sup>b</sup> Proportion of dominant genes
- <sup>c</sup> Balance of positive and negative alleles
- d Heritability, narrow sense
- e Heritability, broad sense

The graph of Vr and Wr for shoot regeneration is shown in Fig. 1. There were no significant differences in the slope of the regression of Wr on Vr from unity, and the homogeneity of Wr-Vr suggested the absence of nonallelic interaction. Cultivars located close to the origin have more dominant alleles, while those far from the origin have more recessive ones. 'Arabella' and 'Doral' carried exclusively dominant alleles, and 'Bridger', 'Cascade' and 'Norin 30' possessed an excess of recessive ones. The analysis of sub-diallels indicated that 'Arabella' and 'Doral' are completely dominant parents and 'Cascade' and 'Norin 30' are completely recessive parents (Fig. 1b). The regression line that intercepted the Wr-axis was above the origin, indicating that incomplete dominance was observed. The correlation coefficient between Vr+Wr and the parental value was r=-0.919, suggesting that the action of dominant genes was positive for shoot regeneration. The same results were found in regeneration from the seed-derived callus of rice (Abe and Futsuhara 1991; Taguchi-Shiobara et al 1997). In contrast, a negative effect of dominant genes for regeneration was reported in the regeneration from the leaf disc of tomato (Frankenberger et al. 1981) and anther culture (Quimio and Zapata 1990), and a seed-derived cell suspension culture (Tsukahara et al. 1995) of rice. This may be due to differences in the materials employed, the culture conditions and the explant tissues.

The genetic components of variation and the genetic information estimated from the sub-diallel table are presented in Table 4. Additive genetic variance (D) was larger than the dominance genetic variances (H1 and H2). The mean degree of dominance was 0.759, indicating incomplete dominance. This confirmed the results of the graphical analysis as shown in Fig. 1. The broad- and narrow-sense heritabilities were 0.973 and 0.819, respectively. Such a high narrow-sense heritability is considered to be due to the major contribution of additive gene effects to genetic variation in shoot regeneration ability.

The inheritance of in vitro traits have been mainly studied in cereal crops such as rice, barley, wheat and maize. To our knowledge, this is the first report clarifying genetic factors for regeneration ability in *Brassica* crops. The present results indicated that the ability of shoot regeneration from a cotyledonary explant is mainly controlled by dominant nuclear genes, and that such genes can be accumulated into agronomically important cultivars. To identify the molecular makers linked with the genes related to shoot regeneration ability, the production of recombinant imbred lines between 'Arabella' and 'Norin 30' is currently in progress.

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### References

Abe T, Futsuhara Y (1991) Diallel analysis of callus growth and plant regeneration in rice seed-callus. Jpn J Genet 66:129–140

Armstrong CL, Romeo-Severson J, Hodges TK (1992) Improved tissue culture response of an elite maize imbred though backcross breeding, and identification of chromosomal regions important for regeneration by RFLP analysis. Theor Appl Genet 84:755–762

Babic V, Datla RS, Scoles GJ, Keller WA (1998) Development of an efficient *Agrobacterium*-mediated transformation system for *Brassica carinata*. Plant Cell Rep 17:183–188

Frankenberger, EA, Hasegawa PM, Tigchelaar C (1981) Diallel analysis of shoot-forming capacity among selected tomato genotypes. Z Pflanzenphysiol 102:233–242

Hayman B (1954a) The analysis of variance of diallel tables. Biometrics 10:235–244

Hayman B (1954b) The theory and analysis of diallel crosses. Genetics 39:789–809

Hachey JE, Sharma KK, Moloney MM (1991) Efficient shoot regeneration of *Brassica campestris* using cotyledon explants cultured in vitro. Plant Cell Rep 9:549–554

Jinks JL (1954) The analysis of continuous variation in a diallel cross of *Nicotiana rustica* varieties. Genetics 39:767–787

Komatsuda T, Enomoto S, Nakajima, K (1989) Genetics of callus proliferation and shoot differentiation in barley. J Hered 80: 345–350

Komatsuda T, Taguchi-Shiobara F, Oka S, Takaiwa F, Annaka T, Jacobsen H-J (1995) Transfer and mapping of the shoot-differentiation locus *Shd1* in barley chromosome 2. Genome 38: 1009–1014

Koornneef M, Bade J, Hanhart C, Horsman K, Schel J, Soppe W, Verkerk R, Zabel P (1993) Characterization and mapping of a gene controlling shoot regeneration in tomato. Plant J 3:131–141

Lazar MD, Baenziger PS, Schaeffer GW (1984) Combining abilities and heritability of callus formation and plantlet regeneration in wheat (*Triticum aestivum* L.) anther cultures. Theor Appl Genet 68:131–134

Mather K, Jinks JL (1971) Biometrical genetics. Chapman and Hall, London

Moloney MM, Walker JM, Sharma KK (1989) High-efficiency transformation of *Brassica napus* using *Agrobacterium* vectors. Plant Cell Rep 8:238–242

Murashige T, Skoog F (1962) A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol Plant 15: 473–497

Narasimhulu SB, Chopra VL (1988) Species-specific shoot regeneration response of cotyledonary explants of Brassicas. Plant Cell Rep 7:104–106

- Ono Y, Takahata Y, Kaizuma N (1994) Effect of genotype on shoot regeneration from cotyledonary explants of rapeseed (*Brassica napus* L.) Plant Cell Rep 14:13–17
- Palmer CE, Keller WA (1994) In vitro culture of oilseeds. In: Vasil IK, Thrope TA (eds) Plant tissue culture. Kluwer Academic Publishers, Dordrecht, pp 413–455
- Peng J, Hodges T (1989) Genetic analysis of plant regeneration in rice (*Oryza sativa* L.). In Vitro Cell Dev Biol 25:91–94
- Petolino JF, Thompson SA (1987) Genetic analysis of anther culture response in maize. Theor Appl Genet 74:284–286
- Quimio CA, Zapata FJ (1990) Diallel analysis of callus induction and green-plant regeneration in rice anther culture. Crop Sci 30:188–192
- Taguchi-Shiobara F, Komatsuda T, Oka S (1997) Comparison of two indices for evaluating regeneration ability in rice (*Oryza sativa* L.) though a diallel analysis. Theor Appl Genet 94:378–382

- Tsukahara M, Hirosawa T, Nagai E, Kato H, Ikeda R, Maruyama K (1995) Genetic analysis of plant regeneration ability in cell-suspension cultures of rice (*Oryza sativa* L.). Breed Sci 45: 425–428
- Ukai Y (1989) A microcomputer program DIALL for diallel analysis of quantitative characters. Jpn J Breed 39:107–109
- Yamagishi M, Yano M, Fukuta Y, Fukui K, Otani M, Shimada T (1996) Distorted segregation of RFLP markers in regenerated plants derived from anther culture of F<sub>1</sub> hybrids of rice. Genes Genet Syst 71:37–41
- Zhang F-L, Takahata Y, Xu J-B (1998) Medium and genotype factors influencing shoot regeneration from cotyledonary explants of Chinese cabbage (*Brassica campestris* L. ssp. *pekinensis*). Plant Cell Rep 17:780–786